NOTES

BIOSYNTHESIS OF AURODOX (ANTIBIOTIC X-5108).

INCORPORATION OF ¹⁴C-LABELLED PRECURSORS INTO AURODOX^{1,2)}

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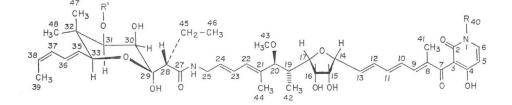
Streptomyces goldiniensis produces aurodox³⁾ (1) and small amounts of its N-demethyl homolog mocimycin (2)⁴⁾ which is probably identical⁵¹ with kirromycin^{6,7)}. These antibiotics and a recently discovered disaccharide derivative of aurodox (3)⁸⁾ are primarily active against Grampositive bacteria and enhance the growth of farm animals. The chemical structures, including stereochemistry, have been elucidated for aurodox and mocimycin^{5,9)}. Since aurodox is a member of a new class of antibiotics, biosynthetic studies of aurodox were of particular interest. In this report we describe the incorporation of ¹⁴Clabelled precursors into aurodox.

The carbon skeleton of aurodox suggests that the antibiotic is in part derived from coupling of acetate units *via* the polyketide biosynthetic route¹⁰. The C-methyl groups could arise through transmethylations from suitable C_1 donors¹¹ or by the insertion of propionate¹² or isoprenoid units. The C-ethyl group could be generated by the incorporation of a butyrate unit into the polyketide chain¹³, whereas the pyridone ring of aurodox could be derived *via* the nicotinic acid pathway¹⁴¹ or, alternatively, from cadaverine. These possibilities were examined experimentally by measuring the incorporation of various ¹⁴C-labelled precursors into aurodox (Table 1).

Radioactively labelled substrates at $10\sim100$ µCi per 100 ml broth were individually added to fermentation cultures of *S. goldiniensis* 2 days after inoculation. After a further 5-day incubation period, aurodox was isolated according to procedures previously described³⁾.

According to the data in Table 1 the labels of methionine, glycine, acetate, propionate, butyrate, and β -hydroxybutyrate were incorporated into aurodox. The pattern of these incorporations (Table 2) was determined by measuring the radioactivity in fragments of aurodox derived by the degradation scheme^{15,161} shown in Fig. 1.

The distribution of radioactivity in acetatederived aurodox reveals little about the assembly of the polyketide chain. In contrast, the incorporation patterns of propionate and butyrate clearly show that an intact unit of each of these precursors is incorporated into aurodox. Localization of all three propionate carbons in fragment 8 suggests that propionate is incorporated into C-41 and C-8 as well as either C-7 or C-9. Butyrate labelling of goldinono-1,4-lactone-3,7hemiacetal (5) may be due to incorporation into the C-ethyl carbons 45 and 46, and the two adjacent carbons 27 and 28. β -Hydroxybutyrate, a known precursor of butyrate, was also incorporated primarily into 5. Perhaps the most interesting result is the extent of incorporation of methionine-methyl-14C (9.7%) and the locali-



- I Aurodox (X-5108) $R = CH_3$; R' = H
- 2 Mocimycin (Kirromycin) R = R'= H
- 3 Efrotomycin $R = CH_3$; $R' = Disaccharide (C_{15}H_{27}O_8)$

zation of radioactivity in **5** (33%). These observations suggest that the geminal C-methyl groups at C-32 are derived from methioninemethyl; the lack of any incorporation of mevalonate-2-¹⁴C and of incorporation of propionate into **5** are consistent with this proposal. The incorporation pattern of methionine methyl-¹⁴C also suggests that the N-methyl (C-40), O-methyl (C-43), and two of the three C-methyls, (C-42 and

Table 1. Incorporation of ¹⁴C-labelled substrates into aurodox

Substrate (sodium salts)	Percentage incorporation	
Acetate-1-14C	2.6	
Acetate-2-14C	2.1	
Propionate-1-14C	2.2	
Propionate-2-14C	1.6	
Propionate-3-14C	0.9	
Butyrate-1-14C	2.2	
Butyrate-2-14C	2.6	
Butyrate-3,4-14C	2.2	
β -Hydroxybutyrate-3- ¹⁴ C	2.5	
Methionine-methyl-14C	9.7	
Glycine-1-14C	0.8	
Glycine-2-14C	2.9	
Cadaverine-1,5-14C	0.6	
Mevalonate-2-14C	0	
Formate-14C	0	
Nicotinic acid-7-14C	0	
Quinolinic acid-6-14C	0	

C-44) are also derived from methionine. Glycine-2-¹⁴C incorporation follows the same patterns as methionine-methyl-¹⁴C, presumably because the methylene carbon of glycine serves as a C_1 -donor.

The origin of the pyridone moiety of aurodox remains to be determined. The nicotinic acid pathway is apparently not involved since nicotinic acid- 7^{-14} C and quinolinic acid- 6^{-14} C were not incorporated. The possibility that cadaverine plays a role in the synthesis of the pyridone group was ruled out by the observation that the total radioactivity in the goldinamine part isolated as **6**, was lower than that contained in the goldinonic acid portion, isolated as **5**.

From these results, the following conclusions can be drawn: (1) Only one propionate unit and one butyrate unit are utilized by *S. goldiniensis* in the assembly of the antibiotic carbon skeleton; (2) six methyl groups are most likely derived *via* transfer of one-carbon units; (3) the pyridone group is not derived from nicotinic acid or cadaverine. Spectroscopic studies with ¹³Cenriched aurodox derived from ¹³C-labelled precursors should permit more specific determination of the biosynthetic origin and are now in progress.

References and notes

 A preliminary account of this study was presented at the Annual Meeting of the American Society for Microbiology, 1974. (Abstr. Ann. Meeting American Society for Microbiology,

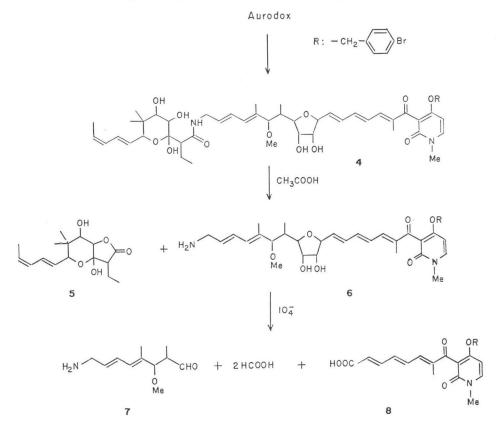
Table 2. Distribution of label radioactivity in aurodox derived from various ¹⁴C-labelled precursors

Labelled precursor	Molar radioactivity, percent of aurodox			
	5	6	7	8
Acetate-1-14C	49.6	50.5	38.9	11.5
Acetate-2-14C	42.6	57.4	41.4	16.0
Propionate-1-14C	1.0	98.7	1.2	89.1
Propionate-2-14C	5.1	99.2	4.5	87.6
Propionate-3-14C	5.7	92.7	5.4	88.4
Butyrate-1- ¹⁴ C	73.8	29.5	3.5	22.5
Butyrate-2-14C	73.2	37.4	12.1	23.3
Butyrate-3,4-14C	62.7	44.1	8.5	33.8
β -Hydroxybutyrate-3- ¹⁴ C	79.1	20.9	ND*	ND
Methionine-methyl-14C	33.8	67.3	41.4	26.0
Glycine-2-14C	32.4	67.2	52.8	12.0
Cadaverine-1,5-14C	63.0	37.0	ND	ND

* ND=not determined

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Fig. 1. Degradation of aurodox bromobenzyl ether



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